

FORM-PTO-1390
(Rev. 12-29-99)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

001560-396

U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.5)

To be assigned

09/807541

INTERNATIONAL APPLICATION NO.
PCT/JP00/05425INTERNATIONAL FILING DATE
11 August 2000PRIORITY DATE CLAIMED
13 August 1999

TITLE OF INVENTION

MICROORGANISMS THAT EXTRACELLULARLY SECRETE LIPIDS AND METHODS OF PRODUCING LIPID AND LIPID VESICLES ENCAPSULATING LIPIDS USING SAID MICROORGANISMS

APPLICANT(S) FOR DO/EO/US

Kengo AKIMOTO, Hiroshi KAWASHIMA, and Sakayu SHIMIZU

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and the PCT Articles 22 and 39(1).
4. ☐ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern other document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:

PCT Notice Informing Applicant of the Communication of the International Application to the Designated Offices (Form PCT/IB/308)

Cover page from published PCT application (WO 01/12780)

JP PCT Request Form

Translations of Receipt in the Case of an Original Deposit (dated Dec. 22, 1986 and Aug. 11, 2000)

U.S. APPLICATION NO. (if known / see 37 CFR 1.50) To be assigned		INTERNATIONAL APPLICATION NO. PCT/JP00/05425		ATTORNEY'S DOCKET NUMBER 001560-396	
17. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS	PTO USE ONLY
Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1,000.00 (960) International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00 (970) International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00 (958) International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00 (956) International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00 (962) ENTER APPROPRIATE BASIC FEE AMOUNT =				\$ 860.00	
Surcharge of \$130.00 (154) for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492(e)). 20 <input type="checkbox"/> 30 <input type="checkbox"/>				\$	
Claims	Number Filed	Number Extra	Rate		
Total Claims	79 -20 =	59	X\$18.00 (966)	\$ 1,062.00	
Independent Claims	9 -3 =	6	X\$80.00 (964)	\$ 480.00	
Multiple dependent claim(s) (if applicable)			+ \$270.00 (968)	\$	
TOTAL OF ABOVE CALCULATIONS =				\$ 2,402.00	
Reduction for 1/2 for filing by small entity, if applicable (see below).				\$	
SUBTOTAL =				\$ 2,402.00	
Processing fee of \$130.00 (156) for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492(f)). 20 <input type="checkbox"/> 30 <input type="checkbox"/>				\$	
TOTAL NATIONAL FEE =				\$ 2,402.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 (581) per property +				\$ 40.00	
TOTAL FEES ENCLOSED =				\$ 2,442.00	
				Amount to be: refunded \$	
				charged \$	
a. <input type="checkbox"/> Small entity status is hereby claimed.					
b. <input type="checkbox"/> A check in the amount of \$_____ to cover the above fees is enclosed.					
c. <input checked="" type="checkbox"/> Please charge my Deposit Account No. <u>02-4800</u> in the amount of \$ <u>2,442.00</u> to cover the above fees. A duplicate copy of this sheet is enclosed.					
d. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>02-4800</u> . A duplicate copy of this sheet is enclosed.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO: Ronald L. Grudziecki BURNS, DOANE, SWECKER & MATHIS, L.L.P. P.O. Box 1404 Alexandria, Virginia 22313-1404 (703) 836-6620					
				SIGNATURE Donna M. Meuth NAME	
April 13, 2001				36,607 REGISTRATION NUMBER	

Patent
Attorney's Docket No. 001560-396

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of)	
)	
Kengo AKIMOTO et al)	Group Art Unit: To be assigned
)	
Application No.: To be assigned)	Examiner: To be assigned
(National Phase of PCT Int'l Appln. No.)	
PCT/JP00/05425 filed August 11, 2000))	
)	
Filed: April 13, 2001)	
)	
For: MICROORGANISMS THAT)	
EXTRACELLULARLY SECRETE)	
LIPIDS AND METHODS OF)	
PRODUCING LIPID AND LIPID)	
VESICLES ENCAPSULATING LIPIDS)	
USING SAID MICROORGANISMS)	

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

Prior to examination of the above-identified application on the merits, please amend
the application as follows:

IN THE CLAIMS

Please delete claim 44 without prejudice or disclaimer

Please replace claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, 45 and 49-51 as
follows.

3. (Amended) The microorganism according to claim 1, which is filamentous
fungus.

7. (Amended) The microorganism according to claim 1, which has a property
of forming lipid vesicles containing a lipid around the colonies when said microorganism is

grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

8. (Amended) The microorganism according to claim 1, which is selected by artificially treating a microorganism having an ability to accumulate an unsaturated fatty acid-containing lipid in the cell.

9. (Amended) The microorganism according to claim 1, which is selected by artificially treating a microorganism having an ability of accumulating an unsaturated fatty acid-containing lipid in the cell, by culturing the obtained strains on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and then by selecting those strains that make the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

10. (Amended) The microorganism according to claim 1, which can be turned into a spheroplast or a protoplast.

12. (Amended) The microorganism according to claim 1, wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

13. (Amended) The microorganism according to claim 1, wherein said unsaturated fatty acids are arachidonic acid.

17. (Amended) The lipid vesicles according to claim 15, wherein said lipid vesicles are produced by a microorganism.

18. (Amended) Lipid vesicles encapsulating a lipid obtained from a culture liquid prepared by culturing the microorganism according to claim 1 in a liquid medium.

19. (Amended) The lipid vesicles according to claim 15, which can be uniformly dispersed in water or a hydrophilic substance.
20. (Amended) The lipid vesicles according to claim 15, which stably retains the lipid encapsulated within said lipid vesicles against oxidation.
21. (Amended) The lipid vesicles according to claim 15, which can be separated by centrifugation.
22. (Amended) The lipid vesicles according to claim 15, wherein the membrane of said lipid vesicles comprises sugar, protein, and lipid.
23. (Amended) The lipid vesicles according to claim 15, which has an average diameter of 0.2 to 10 μm .
24. (Amended) The lipid vesicles according to claim 15, wherein the lipid encapsulated in said lipid vesicles is a lipid in which 50% or more is triglyceride.
25. (Amended) A lipid isolated from the lipid vesicles according to claim 15.
26. (Amended) A food, a cosmetic, or an animal feed comprising the lipid vesicles according to claim 15 added thereto.
30. (Amended) A method of producing lipid vesicles which method comprises culturing the microorganism according to claim 1 in a liquid medium and then collecting the lipid vesicles encapsulating a lipid from the culture liquid.
31. (Amended) A method of producing lipid vesicles which method comprises continuously culturing the microorganism according to claim 1 in a liquid medium and then continuously collecting the lipid vesicles encapsulating a lipid from the culture liquid.

32. (Amended) A method of producing a lipid which method comprises culturing the microorganism according to claim 1 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, and separating a lipid containing fatty acids from said lipid vesicles.

33. (Amended) A method of producing unsaturated fatty acids which method comprises culturing the microorganism according to claim 1 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, separating the lipid containing fatty acids from said lipid vesicles, and isolating the unsaturated fatty acids from said lipid.

36. (Amended) The microorganism according to claim 34, which is a filamentous fungus.

37. (Amended) The microorganism according to claim 34, which has a property of forming lipid-containing lipid vesicles around the colonies thereof when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

38. (Amended) The microorganism according to claim 34 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

39. (Amended) The microorganism according to claims 35 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, and by selecting, from the strains obtained,

strains that make the culture liquid cloudy and then separates a lipid layer when cultured in a transparent liquid medium.

40. (Amended) The microorganism according to claim 34, which can be turned into a spheroplast or a protoplast.

41. (Amended) The microorganism according to claim 34, wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

42. (Amended) A method of producing a lipid containing unsaturated fatty acids which method comprises culturing the microorganism according to claim 34 in a liquid medium and collecting the lipid from the culture liquid.

43. (Amended) A method of producing a lipid containing unsaturated fatty acids which method comprises continuously culturing the microorganism according to claim 34 in a liquid medium and then continuously collecting the lipid from the culture liquid.

45. (Amended) The screening method according to claim 74 wherein said unsaturated fatty acids have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

46. (Amended) The screening method according to claim 74 wherein said microorganism is a filamentous fungus.

47. (Amended) A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, and culturing the strains obtained on a solid medium to

determine strains of which colonies are covered with lipid-containing lipid vesicles at the periphery.

48. (Amended) A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, [by] culturing the strains obtained on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and further culturing the selected strains in a transparent liquid medium to determine strains for which the culture liquid becomes cloudy.

49. (Amended) The screening method according to claim 47, wherein said artificial manipulation is mutation treatment with N-methyl-N'-nitro-N-nitrosoguanidine (NTG).

50. (Amended) The screening method according to claim 47, wherein said artificial manipulation is mutation treatment, gene manipulation, or cell fusion.

51. (Amended) A microorganism selected by the screening method according to claim 74.

Please add new claims 52-80 as follows:

--52. The microorganism according to claim 34, wherein said unsaturated fatty acid is selected from the group consisting of γ -linolenic acid, arachidonic acid, DHA and ω 9 highly unsaturated fatty acids.

53. The microorganisms according to claim 8, wherein said artificial treatment is selected from the group consisting of mutation, gene manipulation and cell fusion.

54. The microorganism according to claim 1, wherein at least one of a reaction in the microorganism selected from the group consisting of $\Delta 5$ desaturation reaction, $\Delta 6$ desaturation reaction, $\Delta 9$ desaturation reaction, $\Delta 12$ desaturation reaction, $\omega 3$ desaturation reaction and chain elongation reaction is enhanced, or reduced or missing.

55. The microorganism according to claim 6, which has a property of forming lipid vesicles containing a lipid around the colonies when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

56. The microorganism according to claim 7, which is selected by artificially treating a microorganism having an ability to accumulate an unsaturated fatty acid-containing lipid in the cell.

57. The microorganism according to claim 8, which is selected by artificially treating a microorganism having an ability of accumulating an unsaturated fatty acid-containing lipid in the cell, by culturing the obtained strains on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and then by selecting those strains that make the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

58. The microorganism according to claim 12, wherein said unsaturated fatty acids are arachidonic acid.

59. The lipid vesicles according to claim 15 wherein said unsaturated fatty acids are unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

60. Lipid vesicles encapsulating a lipid obtained from a culture liquid prepared by culturing the microorganism according to claim 55 in a liquid medium.

61. The lipid vesicles according to claim 60, wherein the lipid encapsulated in said lipid vesicles is a lipid in which 50% or more is triglyceride.

62. A lipid isolated from the lipid vesicles according to claim 62.

63. A food, a cosmetic, or an animal feed comprising the lipid vesicles according to claim 61 added thereto.

64. A food, a cosmetic, a pharmaceutical or an animal feed comprising the lipid according to claim 62 added thereto.

65. A method of producing lipid vesicles which method comprises culturing the microorganism according to claim 57 in a liquid medium and then collecting the lipid vesicles encapsulating a lipid from the culture liquid.

66. A method of producing lipid vesicles which method comprises continuously culturing the microorganism according to claim 57 in a liquid medium and then continuously collecting the lipid vesicles encapsulating a lipid from the culture liquid.

67. A method of producing a lipid which method comprises culturing the microorganism according to claim 57 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, and separating a lipid containing fatty acids from said lipid vesicles.

68. A method of producing unsaturated fatty acids which method comprises culturing the microorganism according to claim 56 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, separating the lipid containing fatty acids from said lipid vesicles, and isolating the unsaturated fatty acids from said lipid.

69. The microorganism according to claim 35, which is a filamentous fungus.

70. The microorganism according to claim 37 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

71. The microorganism according to claim 39, wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

72. A method of producing a lipid containing unsaturated fatty acids which method comprises culturing the microorganism according to claim 39 in a liquid medium and collecting the lipid from the culture liquid.

73. A method of producing a lipid containing unsaturated fatty acids which method comprises continuously culturing the microorganism according to claim 39 in a liquid medium and then continuously collecting the lipid from the culture liquid.

74. A screening method for determining whether a microorganism has an ability of extracellularly secreting a lipid containing unsaturated fatty acids comprising culturing a microorganism in a transparent liquid medium, and determining whether the culture liquid becomes cloudy.

75. A screening method according to claim 74, wherein the microorganism is selected from genus *Mortierella*.
76. A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, and by culturing the strains obtained on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery.
77. The screening method according to claim 48, wherein said artificial manipulation is mutation treatment with N-methyl-N'-nitro-N-nitrosoguanidine (NTG).
78. The screening method according to claim 48, wherein said artificial manipulation is mutation treatment, gene manipulation, or cell fusion.
79. A microorganism selected by the screening method according to claim 47.
80. A microorganism selected by the screening method according to claim 48.--

REMARKS

Prior to examination, entry of the foregoing is respectfully requested.

Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, 45, 46 and 49-51 have been amended simply to delete multiple dependencies in the claims and correct claim dependencies.

Minor amendments relating to matters of form only have also been made.

Claim 44 has been deleted and rewritten as new claim 74.

New claims 52-73 and 75-80 have been added, directed to preferred embodiments of the invention in view of the deletion of multiple dependent claims. Support for these claims may be found at the very least at pages 9-11 of the specification and in original claims 1-51.

In the event that there are any questions relating to this Preliminary Amendment, or to the application in general, it would be appreciated if the Examiner would telephone the undersigned attorney at (508) 339-3684 concerning such questions so that prosecution of this application may be expedited.

Early and favorable action in the form of a Notice of Allowance is respectfully requested and believed to be in order.

Respectfully submitted,

BURNS, DOANE, SWECKER & MATHIS, L.L.P.

By: 

Donna M. Meuth

Registration No. 36,607

P.O. Box 1404
Alexandria, Virginia 22313-1404
(703) 836-6620

Date: April 13, 2001

Attachment to Preliminary Amendment dated April 13, 2001

Marked-up Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, and 49-51

3. (Amended) The microorganism according to claim 1 [or 2], which is filamentous fungus.

7. (Amended) The microorganism according to [any one of claims 1 to 6] claim 1, which has a property of forming lipid vesicles containing a lipid around the colonies when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

8. (Amended) The microorganism according to [any one of claims 1 to 7] claim 1, which is selected by artificially treating a microorganism having an ability to accumulate an unsaturated fatty acid-containing lipid in the cell.

9. (Amended) The microorganism according to [any one of claims 1 to 7] claim 1, which is selected by artificially treating a microorganism having an ability of accumulating an unsaturated fatty acid-containing lipid in the cell, by culturing the obtained strains on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and then by selecting those strains that make the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

10. (Amended) The microorganism according to [any one of claims 1 to 9] claim 1, which can be turned into a spheroplast or a protoplast.

12. (Amended) The microorganism according to [any one of claims 1 to 11] claim 1, wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

13. (Amended) The microorganism according to [any one of claims 1 to 12] claim 1, wherein said unsaturated fatty acids are arachidonic acid.

17. (Amended) The lipid vesicles according to [claim 15 or 16] claim 15, wherein said lipid vesicles are produced by a microorganism.

18. (Amended) Lipid vesicles encapsulating a lipid obtained from a culture liquid prepared by culturing the microorganism according to [any one of claims 1 to 14] claim 1 in a liquid medium.

Attachment to Preliminary Amendment dated April 13, 2001

Marked-up Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, and 49-51

19. (Amended) The lipid vesicles according to [any one of claims 15 to 18] claim 15, which can be uniformly dispersed in water or a hydrophilic substance.
20. (Amended) The lipid vesicles according to [any one of claims 15 to 18] claim 15, which stably retains the lipid encapsulated within said lipid vesicles against oxidation.
21. (Amended) The lipid vesicles according to [any one of claims 15 to 18] claim 15, which can be separated by centrifugation.
22. (Amended) The lipid vesicles according to [any one of claims 15 to 21] claim 15, wherein the membrane of said lipid vesicles comprises sugar, protein, and lipid.
23. (Amended) The lipid vesicles according to [any one of claims 15 to 22] claim 15, which has an average diameter of 0.2 to 10 μm .
24. (Amended) The lipid vesicles according to [any one of claims 15 to 23] claim 15, wherein the lipid encapsulated in said lipid vesicles is a lipid in which 50% or more is triglyceride.
25. (Amended) A lipid isolated from the lipid vesicles according to [any one of claims 15 to 24] claim 15.
26. (Amended) A food, a cosmetic, or an animal feed comprising the lipid vesicles according to [any one of claims 15 to 24] claim 15 added thereto.
30. (Amended) A method of producing lipid vesicles which method comprises culturing the microorganism according to [any one of claims 1 to 14] claim 1 in a liquid medium and then collecting the lipid vesicles encapsulating a lipid from the culture liquid.
31. (Amended) A method of producing lipid vesicles which method comprises continuously culturing the microorganism according to [any one of claims 1 to 14] claim 1 in a liquid medium and then continuously collecting the lipid vesicles encapsulating a lipid from the culture liquid.
32. (Amended) A method of producing a lipid which method comprises culturing the microorganism according to [any one of claims 1 to 14] claim 1 in a liquid

Attachment to Preliminary Amendment dated April 13, 2001

Marked-up Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, and 49-51

medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, and separating a lipid containing fatty acids from said lipid vesicles.

33. (Amended) A method of producing unsaturated fatty acids which method comprises culturing the microorganism according to [any one of claims 1 to 14] claim 1 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, separating the lipid containing fatty acids from said lipid vesicles, and isolating the unsaturated fatty acids from said lipid.

36. (Amended) The microorganism according to claim 34, [or 35] which is a filamentous fungus.

37. (Amended) The microorganism according to [any one of claims 34 to 36] claim 34, which has a property of forming lipid-containing lipid vesicles around the colonies thereof when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

38. (Amended) The microorganism according to [any one of claims 34 to 37] claim 34 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

39. (Amended) The microorganism according to claims 35 [or 36] obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, and by selecting, from the strains obtained, strains that make the culture liquid cloudy and then separates a lipid layer when cultured in a transparent liquid medium.

40. (Amended) The microorganism according to [any one of claims 34 to 39] claim 34, which can be turned into a spheroplast or a protoplast.

Attachment to Preliminary Amendment dated April 13, 2001

Marked-up Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, and 49-51

41. (Amended) The microorganism according to [any one of claims 34 to 40] claim 34, wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

42. (Amended) A method of producing a lipid containing unsaturated fatty acids which method comprises culturing the microorganism according to [any one of claims 34 to 41] claim 34 in a liquid medium and collecting the lipid from the culture liquid.

43. (Amended) A method of producing a lipid containing unsaturated fatty acids which method comprises continuously culturing the microorganism according to [any one of claims 34 to 41] claim 34 in a liquid medium and then continuously collecting the lipid from the culture liquid.

45. (Amended) The screening method according to claim [44] 74 wherein said unsaturated fatty acids have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

46. (Amended) The screening method according to claim [44] 74 wherein said microorganism is a filamentous fungus.

47. (Amended) A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, and [by] culturing the strains obtained on a solid medium to [select] determine strains of which colonies are covered with lipid-containing lipid vesicles at the periphery.

48. (Amended) A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, [by] culturing the strains obtained on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery,

Attachment to Preliminary Amendment dated April 13, 2001

Marked-up Claims 3, 7-10, 12, 13, 17-26, 30-33, 36-43, and 49-51

and [by] further culturing the selected strains in a transparent liquid medium to [select] determine strains for which the culture liquid becomes cloudy.

49. (Amended) The screening method according to claim 47, [or 48] wherein said artificial manipulation is mutation treatment with N-methyl-N'-nitro-N-nitrosoguanidine (NTG).

50. (Amended) The screening method according to claim 47, [or 48] wherein said artificial manipulation is mutation treatment, gene manipulation, or cell fusion.

51. (Amended) A microorganism selected by the screening method according to [any one of claims 44 to 50] claim 74.

- 1 -

DESCRIPTION

5 MICROORGANISMS THAT EXTRACELLULARLY SECRETE LIPIDS
 AND METHODS OF PRODUCING LIPID AND LIPID VESICLES
 ENCAPSULATING LIPIDS USING SAID MICROORGANISMS

Field of the Invention

10 The present invention relates to microorganisms
 having a property of extracellularly secreting a lipid
 containing unsaturated fatty acids encapsulated in small
 vesicles, or to microorganisms having a property of
 extracellularly secreting a lipid containing unsaturated
 fatty acids that have 18 carbons and three or more double
 bonds or 20 or more carbons and two or more double bonds,
15 and more specifically to microorganisms having a property
 of extracellularly secreting a lipid containing
 unsaturated fatty acids encapsulated in small vesicles,
 said microorganisms being obtained by artificially
 treating microorganisms that have an ability of producing
20 and intracellularly accumulating a lipid containing
 unsaturated fatty acids, or to microorganisms having a
 property of extracellularly secreting a lipid containing
 unsaturated fatty acids that have 18 carbons and three or
 more double bonds or 20 or more carbons and two or more
25 double bonds, said microorganisms being obtained by
 artificially manipulating microorganisms that have an
 ability of producing and intracellularly accumulating a
 lipid containing unsaturated fatty acids that have 18
 carbons and three or more double bonds or 20 or more
30 carbons and two or more double bonds, and to methods of
 efficiently producing a lipid containing unsaturated
 fatty acids using said microorganisms, as well as to
 methods of screening said microorganisms, and furthermore
 to lipid vesicles encapsulating a lipid that contains
35 unsaturated fatty acids, and to foods, cosmetics, and
 animal feeds comprising said lipid vesicles added
 thereto.

Background Art

In recent years, various biological activities of highly unsaturated fatty acids have attracted attention. For example, arachidonic acid is considered to be a precursor of prostaglandins, thromboxanes, prostacyclins, leucotrienes and the like that have biological activities such as uterine contraction and relaxation, vasodilation, and hypotensive activity. These substances have been under intensive research, together with docosahexaenoic acid, in recent years, as elements essential for the development of babies and infants. Various foods, cosmetics, and animal feeds to which a lipid containing highly unsaturated fatty acids such as γ -linolenic acid, dihomogamma-linolenic acid, and eicosapentaenoic acid in addition to arachidonic acid and docosahexaenoic acid have been added are also attracting attention, and some of the products to which highly unsaturated fatty acids has been added are commercially available.

Accordingly, methods of efficiently producing these highly unsaturated fatty acids also are being studied intensively.

For example, methods of efficiently producing arachidonic acid, dihomogamma-linolenic acid, γ -linolenic acid, or eicosapentaenoic acid by fermentation have been developed using microorganisms belonging to, for example, the genus *Mortierella*, specifically the subgenus *Mortierella*, that are known to produce highly unsaturated fatty acids such as arachidonic acid, dihomogamma-linolenic acid, γ -linolenic acid, and eicosapentaenoic acid (Japanese Unexamined Patent Publication (Kokai) No. 63-44891, Japanese Unexamined Patent Publication (Kokai) No. 63-12290, Japanese Unexamined Patent Publication (Kokai) No. 63-14696, Japanese Unexamined Patent Publication (Kokai) No. 5-91887, and Japanese Unexamined Patent

Publication (Kokai) No. 63-14697). There is also known a method of producing mead acid using a mutant strain in which $\Delta 12$ desaturation activity is decreased or missing, said strain being obtained by effecting mutation to microorganisms belonging to genus *Mortierella* subgenus *Mortierella* (Japanese Unexamined Patent Publication (Kokai) No. 5-91888).

Thus, the production of a lipid containing highly unsaturated fatty acids using microorganisms that produce said fatty acids is increasingly becoming a major source of highly unsaturated fatty acids. These microorganisms have a property of not only using highly unsaturated fatty acids they produce as constituents of the cell membrane but also accumulating highly unsaturated fatty acids as fat and oil (triglycerides) containing them as constituent fatty acids in the cell. By utilizing the fats and lipids accumulated in the cell, high productivity of highly unsaturated fatty acids have come to be secured.

In this conventional production method, the amount of fat and oil obtained per culture was a product of the cell mass of the microorganism obtained by culturing and the amount of fat and oil produced per cell, and thereby how to increase the cell mass and the amount of fat and oil per cell was a challenge to be solved in order to attain enhanced production of fat and oil. Research has so far revealed that the selection of culture conditions permits increases in both of the above to a certain extent, but there are certain limits to each of them with increases in the cell mass being limited by physical factors such as the volume of the culture tank and increases in the amount of fat and oil per cell being limited by physiological factors of the microorganism used.

On the other hand, when it is desired to utilize a lipid produced by and accumulated in the cell of the microorganism, it is necessary to collect the cells after

culturing, process the cells with a mill etc., to disrupt the cell membrane, and then to extract the lipid accumulated in the cell.

5 If substances produced by a microorganism could be secreted outside of the cell instead of being accumulated in the cell, the physiological burdens placed on the microorganism by the substances produced can be alleviated and thus the microorganism can continue to produce the products; in the isolation and extraction of microbial products as well, extraction from the culture only is needed, which provides an advantage that treatment becomes simplified and microorganisms can be continuously processed as they remain alive.

10 Based on the above, efforts to extracellularly secret a lipid accumulated in the cell have recently been made by Sakuzo Fukui et al. (BIO INDUSTRY 12: 36-46 (1995)). Sakuzo Fukui et al. conducted research on the production and secretion of lipids by microorganisms to develop novel biological fuel that can replace fossil fuel, and have successfully bred genus *Trichosporon* yeast to convert sugar and n-alkanes to lipids and to secret them outside of the cell. They further demonstrated that the constituent fatty acid species of extracellularly secreted triglyceride (TG) are oleic acid, palmitic acid, linoleic acid, and stearic acid. However, it has a drawback that the TG that is directly secreted outside of the cell is incorporated into the cell again and metabolized.

20 However, microorganisms that have an ability to produce lipids containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, and that have a property of extracellularly secreting the produced lipid, or microorganisms that have an ability of producing an unsaturated fatty acid-containing lipid, and that secrete the produced lipid encapsulated in small vesicles instead of directly secreting them at the time

of extracellular secretion thereof, are not known.

In a study on γ -linolenic acid production by molds described in "Microbially produced fatty acids and their uses", Osamu Suzuki, Fragrance Journal 1989 (6), pp. 67-75, it is reported that a surfactant was added to a culture medium of a microorganism of the genus *Mucor* to allow the leakage of some of lipids outside of the cell. However, it relates to methods of artificially destroying the cell membrane to allow lipids accumulated in the cell to leak out of the cell, and does not utilize the ability of the cell per se to extracellularly secrete lipids produced in the cell.

Thus, there is a need to find microorganisms that have an ability of producing a lipid containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, and have a property of extracellularly secreting the produced lipids, or microorganism that have an ability of producing an unsaturated fatty acid-containing lipid, and that have a property of extracellularly secreting the produced lipids encapsulated in small vesicles, and to develop methods of efficiently producing lipids that contain unsaturated fatty acids using said microorganisms.

Incidentally, in order to find microorganisms that have novel abilities, the development of an efficient screening method for microorganisms having such abilities is a prerequisite. In the above breeding of genus *Trichosporon* yeast that converts sugar and n-alkanes into lipids and extracellularly secrete them, the following screening method was adopted. Thus, yeast colonies that appeared on an agar plate medium (YPD medium etc.) are UV-treated (15 watts at a distance of 30 cm for 15 minutes) (this treatment is intended to suppress the dispersion of colony cells during layering treatment and not to induce mutation), the UV-treated colony plates are layered with a YPD soft agar medium containing 10^5 cells

of a test strains, and are then cultured at 28°C for 2 days. As the test strains, the A-1 strain and the ole-1 strain having auxotrophy for saturated fatty acids and unsaturated fatty acids, respectively, are used and those colonies giving a larger halo (the micro colony ring of the test strain) around them are selected as lipid-secreting strains. The selection of lipid-secreting strains employs two media: a soft agar medium containing or not containing lipase.

10 This method has a major drawback that although it can be applied to screening of yeast for which layering is possible, it cannot be applied to screening of microorganisms for which layering is impossible. There is additional disadvantage that the assay method are
15 complicated.

Thus, in order to find microorganisms that have an ability of producing a lipid containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, said
20 microorganisms having a property of extracellularly secreting the produced lipids, there is a need for the development of a screening method that permits simple and efficient screening of said microorganisms and that can be applied to various microorganisms, and the development
25 of a new screening method for finding microorganisms that can secrete the produced lipids encapsulated in small vesicles at the time of extracellular secretion of the produced lipids.

30 Disclosure of the Invention

Thus, the present invention intends to provide microorganisms that have an ability of producing a lipid containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and
35 two or more double bonds, and have a property of extracellularly secreting the produced lipids, or said microorganisms that have an ability of producing

unsaturated fatty acids, secrete the produced lipids encapsulated in small vesicles, and methods of efficiently producing said lipids using said microorganisms, as well as methods of screening said microorganisms. The present invention also intends to provide lipid vesicles encapsulating a lipid containing unsaturated fatty acids, and to foods, cosmetics, and animal feeds comprising said lipid vesicles added thereto.

After intensive research to attain the above objectives, the inventors of the present invention have found that by artificially treating microorganisms that have an ability of producing and intracellularly accumulating a lipid containing unsaturated fatty acids, it is possible to create microorganisms that have an ability of extracellularly secreting the produced lipids encapsulated in small vesicles.

In order to obtain microorganisms having the desired ability from the group of artificially treated microorganisms or microorganisms collected from nature, the following simple screening method has been found that could be easily practiced by a person skilled in the art. Thus, as a primary screening, artificially treated strains or strains collected from nature are grown on solid media, and thereby those strains providing lipid vesicles around the colonies are selected. Then as a secondary screening, the strains selected in the primary screening are cultured under shaking in a transparent liquid medium (4% glucose, 1% yeast extract, pH 6.0) at 28°C for 2 days. Microorganisms that accumulate lipids in the cell do not make the medium cloudy during culturing, whereas microorganisms that extracellularly secrete the produced lipids make the medium cloudy, and therefore, microorganisms having an ability of extracellularly secreting lipids can be easily screened by a mere visual confirmation of the degree of cloudiness of the culture liquid.

Then, it was found that, the efficient extracellular secretion of the lipid vesicles encapsulating said lipids utilizing the microorganisms obtained may be effected by culturing the microorganisms in a medium having enhanced glucose concentration and/or enhanced pH.

It was also found that the extracellularly secreted lipid vesicles can be easily separated from the culture liquid by centrifugation and that centrifugation and chromatography in addition to extraction with common organic solvents can be used to isolate lipids in the lipid vesicles.

Furthermore, the inventors have found that since the lipid vesicles encapsulating the lipids separated from the culture liquid have a property of being easily dispersed in water or hydrophilic substances and stably retain the lipids against oxidation, the lipid vesicles may be added as they are to foods, cosmetics, or animal feeds thereby to obtain foods, cosmetics, or animal feeds containing unsaturated fatty acids that have novel features, and have completed the present invention.

Embodiment for Carrying Out the Invention

According to the present invention, unsaturated fatty acids that have 18 carbons and three or more double bonds, or 20 or more carbons and two or more double bonds refer to, for example, 5,8,11,14-eicosatetraenoic acid (arachidonic acid), 8,11,14-eicosatrienoic acid (dihomo- γ -linolenic acid), 6,9,12-octadecatrienoic acid (γ -linolenic acid), 5,8,11,14,17-eicosapentaenoic acid, 8,11,14,17-eicosatetraenoic acid, 6,9,12,15-octadecatetraenoic acid (stearidonic acid), 9,12,15-octadecatrienoic acid (α -linolenic acid), 4,7,10,13,16,19-docosahexaenoic acid (DHA), 8,11-eicosadienoic acid, 5,8,11-eicosatrienoic acid (Mead acid), 7,10,13,16-docosatetraenoic acid, 4,7,10,13,16-docosapentaenoic acid, 7,10,13,16,19-docosapentaenoic

acid, and the like.

5 The present invention provides microorganisms that have a property of extracellularly secreting a lipid containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, or microorganisms that have a property of extracellularly secreting a lipid containing unsaturated fatty acids encapsulated in small vesicles. More specifically, the present invention provides 10 microorganisms that can extracellularly secrete a intracellularly produced lipid directly or by encapsulating it in small vesicles, said microorganisms being obtained by artificially treating microorganisms that have an ability of producing and intracellularly accumulating a lipid containing unsaturated fatty acids 15 that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, or that have an ability of producing and intracellularly accumulating a lipid containing unsaturated fatty acids.

20 As used herein, microorganisms that have an ability of producing and intracellularly accumulating a lipid containing unsaturated fatty acids include, for example, conventionally known microorganisms that have an ability of producing γ -linolenic acid or microorganisms that have an ability of producing arachidonic acid, microorganisms 25 that have an ability of producing DHA, microorganisms that have an ability of producing ω 9 highly unsaturated fatty acids, and the like. Microorganisms that have an ability of producing arachidonic acid include 30 microorganisms belonging to genus *Mortierella*, genus *Conidiobolus*, genus *Pythium*, genus *Phytophthora*, genus *Penicillium*, genus *Cladosporium*, genus *Mucor*, genus *Fusarium*, genus *Aspergillus*, genus *Rhodotorula*, genus *Entomophthora*, genus *Echinosporangium* and genus 35 *Saprolegnia*. As microorganisms belonging to genus *Mortierella* subgenus *Mortierella*, there can be mentioned

Mortierella elongata, *Mortierella exigua*, *Mortierella hygrophila*, *Mortierella alpina*, *Mortierella schmuckeri*, *Mortierella minutissima*, and the like. Specifically there can be mentioned strains *Mortierella elongata* IFO8570, *Mortierella exigua* IFO8571, *Mortierella hygrophila* IFO5941, *Mortierella alpina* IFO8568, ATCC16266, ATCC32221, ATCC42430, CBS219.35, CBS224.37, CBS250.53, CBS343.66, CBS527.72, CBS529.72, CBS608.70, and CBS754.68, and the like.

These strains are all available without limitations from the Institute of Fermentation (IFO) in Osaka, Japan, and American Type Culture Collection (ATCC) in the U.S.A., and Centrralbureau voor Schimmelcultures (CBS). It is also possible to use *Mortierella elongata* SAM0219 (FERM P-8703) (FERM BP-1239), a microbial strain isolated from the soil by the present inventor. Microbial strains belonging to these type cultures or those isolated from nature may be used as they are, and spontaneous mutants obtained by effecting growth and/or isolation once or more may be used wherein the mutants have different properties than that of the original strains.

In accordance with the present invention, as microorganisms having the above ability of producing unsaturated fatty acids, there can be used the microorganisms in which at least one of the $\Delta 5$ desaturation reaction, the $\Delta 6$ desaturation reaction, the $\Delta 9$ desaturation reaction, the $\Delta 12$ desaturation reaction, the $\omega 3$ desaturation reaction, and chain elongation reaction is enhanced, or reduced or missing may be obtained by for example, mutation treatment or gene manipulation described below.

As artificial treatment effecting the above microorganisms, there can be mentioned mutation, gene manipulation, cell fusion and the like. Mutation according to the present invention can be conducted by conventional mutation treatments: for example effecting

irradiation treatment (X ray, gamma ray, neutron beams), ultra violet irradiation, and high temperature treatment to induce mutation; and by suspending microorganisms in a suitable buffer etc., to which a mutagen is added followed by incubating for a given time, which is diluted appropriately and inoculated on an agar medium to obtain colonies of mutant strains.

As mutagens, there can be mentioned alkylating agents such as nitrogen mustard, methyl methane sulfonate (MMS), and N-methyl-N'-nitro-N-nitrosoguanidine (NTG), base analogs such as 5-bromouracil, antibiotics such as mitomycin C, base synthesis inhibitors such as 6-mercaptopurine, dyes such as proflavine, certain carcinogens such as 4-nitroquinoline-N-oxide, and other compounds such as manganese chloride and formaldehyde. Microorganisms used may be live cells (mycelia) or spores.

In gene manipulation, conventional gene recombinant technology is used.

From microorganism groups subjected to the above artificial treatment or microorganism groups collected from nature according to conventional methods, strains of interest may be isolated based on the following method. As a primary screening, after strains subjected to artificial treatment or strains collected from nature are plated on a solid medium, the presence of lipid vesicles around colonies are used as an index to select strains, and then as a secondary screening, the strains selected in the primary screening are evaluated in a liquid medium as to whether they secrete a lipid outside of the cell. In an evaluation method, for example, 4 ml of a transparent liquid medium (4% glucose, 1% yeast extract, pH 6.0) is dispensed in a test tube, which is sterilized at 120°C for 20 minutes, and then one platinum loopful of the strain that was selected in the primary screening is inoculated and incubated under shaking at 28°C for 2 days.

Microorganisms like those belonging to genus *Mortierella* subgenus *Mortierella* that produce triglyceride having an unsaturated fatty acid as constituent fatty acids in the cell but do not secrete it outside of the cell, even when cultured in the above liquid medium, do not make the medium cloudy, but when the lipid produced is secreted outside of the cell the medium becomes cloudy, so that the microorganisms that extracellularly secrete the lipid can be easily confirmed. When the cloudiness does not change after being allowed to stand for some time, a lipid has been secreted as lipid vesicles, whereas when the lipid has been directly secreted, a lipid layer rises up to the surface of the medium, so that the two can be easily discriminated from each other. As components of the medium used in the secondary screening, any liquid medium that can become transparent may be selected as appropriate, and any composition of the medium may be selected that is suitable for the growth of the microorganism to be evaluated.

From among the strains selected in this manner, it is preferred to select strains that have an ability to grow and to produce lipids equal to or better than the that of the parent strain used for artificial treatment. Although microorganisms of interest may be selected from either one of the above primary screening or the above secondary screening depending on the microorganism to be selected or the purpose of the study, combination of the two can assure better selection. When the above two screening methods are combined, either the primary screening or the secondary screening may be conducted first.

As a strain obtained by the above method, there can be used *Mortierella alpina* SAM2241 (FERM BP-7272) or SAM 2242 that was derived from *Mortierella alpina* IFO 8568 by the present inventors and that extracellularly secretes an intracellularly produced lipid encapsulated in small

vesicles, but the strain to be used is not limited, and strains that extracellularly secrete the lipid intracellularly produced may be readily obtained by the above screening method, all of which can be used.

5 Microorganisms of the present invention obtained by the above screening method are microorganisms that can be converted to spheroplast or protoplast, and microorganisms of the present invention can be used to
10 obtain protoplast in which the cell wall has been completely removed or spheroplast in which part of the cell wall remains. This property cannot be observed in the parent strain used for artificial treatment such as mutation treatment, and is probably due to the fact that the mutation treatment changed the structure of the cell
15 wall and made it fragile.

 A microbial strain obtained as described above that, for example, strains that extracellularly secretes an intracellularly produced lipid encapsulated in small
20 vesicles in the following method may be used to obtain lipid vesicles or the lipid. In order to culture microorganisms of the present invention, the spores, the mycelia, or the preculture obtained by culturing in advance are inoculated into a liquid medium and cultured.

 In the case of liquid media, the carbon sources used
25 include, but are not limited to, any of glucose, fructose, xylose, saccharose, maltose, soluble starch, molasses, glycerol, mannitol and the like that are commonly used. As nitrogen sources, in addition to natural nitrogen sources such as peptone, yeast extract,
30 malt extract, meat extract, casamino acid, corn steep liquor, soybean protein, defatted soybean, and cottonseed meal, organic nitrogen sources such as urea, and inorganic nitrogen sources such as sodium nitrate, ammonium nitrate, and ammonium sulfate can be used.

35 When desired, inorganic salts such as phosphates, magnesium sulfate, iron sulfate, and copper sulfate, and vitamins can also be used as trace nutrients. The

concentrations of these medium components are not limited as long as they do not adversely affect microbial growth. Generally from the practical viewpoint, carbon sources are in the range of 0.1 to 40% by weight and preferably 1 to 25% by weight. Furthermore, the sequential addition of carbon sources and/or increasing initial concentrations of carbon sources can promote extracellular secretion of lipids. Nitrogen sources may be in the range of 0.1 to 10% by weight and preferably 0.1 to 6% by weight, and the nitrogen sources may be feeded in the middle of culturing.

Though the optimum growth temperature as used herein may vary depending on the microorganism used, it is 5 to 40°C, preferably 20 to 30°C, or after culturing at 20 to 30°C to grow the cell mass, culturing at 5 to 20°C may be continued to produce a lipid containing unsaturated fatty acids. By means of such temperature control, the amount of highly unsaturated fatty acids in the produced fatty acids can be enhanced. pH of the medium is 4 to 10, preferably 5 to 9, and aerated agitation culture, shaking culture, or continuous or stationary culture may be conducted using bioreactors. By using the initial pH of 5 to 9, preferably 6 to 9, and more preferably 7 to 9, extracellular secretion of lipids can be promoted. Culturing is usually conducted for 2 to 30 days, preferably 5 to 20 days, and more preferably 5 to 15 days.

In accordance with the present invention, by adding precursors of the desired unsaturated fatty acids to the medium and then culturing, the production of the desired unsaturated fatty acids, for example 5,8,11,14-eicosatetraenoic acid (arachidonic acid), 8,11,14-eicosatrienoic acid (dihomo- γ -linolenic acid), 6,9,12-octadecatrienoic acid (γ -linolenic acid), 5,8,11,14,17-eicosapentaenoic acid, 8,11,14,17-eicosatetraenoic acid, 6,9,12,15-octadecatetraenoic acid (stearidonic acid),

9,12,15-octadecatrienoic acid (α -linolenic acid), 4,7,10,13,16,19-docosahexaenoic acid (DHA), 8,11-eicosadienoic acid, and 5,8,11-eicosatrienoic acid (Mead acid) can be promoted.

5 Precursors that may be used include, but not limited to, hydrocarbons such as tetradecane, hexadecane, and octadecane, fatty acids such as tetradecanoic acid, hexadecanoic acid, and octadecanoic acid or salts (e.g. sodium salts and potassium salts) or esters thereof, or
10 fat and oil containing fatty acids (e.g. olive oil, coconut oil, palm oil, flaxseed oil, fish oil, and microbial oil) as constituent ingredients.

By adding fat and oil containing, as constituent ingredients unsaturated fatty acids (for example, fish
15 oil and microbial oil), or said unsaturated fatty acids to the medium and culturing, the microorganisms of the present invention incorporate the added unsaturated fatty acids or fat and oil into the cell and extracellularly secrete the lipid containing said unsaturated fatty acids
20 as lipid vesicles. Therefore, even if unsaturated fatty acids that are not originally produced by the microorganism are used, it is possible to produce lipid vesicles encapsulating the lipid containing such unsaturated fatty acids as constituent fatty acids.

25 The total amount of the added substrate containing the above precursor is 0.001 to 10% by weight and preferably 0.5 to 10% by weight relative to the medium. These substrates may be added either before or immediately after inoculating the production
30 microorganism, or after the start of culturing, or they may be added at both time points. The addition after the start of culturing may be once or more than once on an intermittent basis. Alternatively, they may be added continuously. Alternatively, these substrates may be
35 used as the sole carbon source for culturing.

By culturing the microorganisms of the present invention as described above, a lipid containing

unsaturated fatty acids can be produced and accumulated in large quantities in the cell, and said lipid is secreted directly or as lipid vesicles encapsulated in small vesicles. When a liquid medium is used, lipid vesicles or the lipid can be harvested from the culture or the cultured liquid from which cultured cells have been removed as in the following manner.

After culturing is over, cultured cells are separated from the culture using conventional means for separating solids and liquids such as centrifugation and filtration to obtain a medium (referred to as culture liquid) in which lipid vesicles encapsulating lipids are dispersed. From the culture liquid, lipid vesicles encapsulating lipids may be isolated as one containing medium components by lyophilization, or lipid vesicles containing no medium components may only be isolated by conventional centrifugation or column treatment. For example, a centrifuge (TOMYRL-101) or a swing rotor (TS-7) may be used for centrifugation at a maximum centrifugal force of $1000 \times g$ or greater, preferably $1500 \times g$ or greater for about 10 minutes to separate lipid vesicles.

The lipid vesicles obtained in the above methods are composed of sugars, proteins, and lipids. Their composition excluding water comprises 0 to 70%, preferably 20 to 60%, of sugars, 0 to 40%, preferably 10 to 30%, of proteins, and 20 to 100%, preferably 30 to 80%, of lipids. However, the ratio of sugars, proteins, and lipids may vary depending on the culture condition, and is not limited to the above ratio. In fact, water is included in some cases.

The lipid vesicles of the present invention have an average diameter of 0.2 to $10 \mu m$, preferably 2 to $4 \mu m$ with the diameter of the largest lipid vesicle being $40 \mu m$, preferably $10 \mu m$. They can be separated by centrifugation; for example, a centrifuge (TOMYRL-101) or

a swing rotor (TS-7) may be used for centrifugation at a maximum centrifugal force of $1000 \times g$ or greater, preferably $1500 \times g$ or greater, for about 10 minutes to separate lipid vesicles. Furthermore, the lipid vesicles of the present invention may be easily dispersed in water or hydrophilic substances, and they have a property of stably retaining lipids against oxidation.

Harvesting of a lipid in the lipid vesicles can be carried out either directly from the culture liquid before separating the lipid vesicles or from the lipid vesicles separated from the culture liquid. When it is extracted from the lipid vesicles separated from the culture liquid, it is extracted in a manner similar to that conventionally performed to extract lipids from the cells. Thus, lipid vesicles are extracted with an organic solvent under a stream of nitrogen. As organic solvents, ether, hexane, methanol, ethanol, chloroform, dichloromethane, petroleum ether and the like can be used, and satisfactory results can also be obtained by alternate extraction with methanol and petroleum ether, or by extraction with a single layer solvent of chloroform-methanol-water. When they are directly extracted from the culture liquid before separation of the lipid vesicles, similar organic solvents may be used, however, in practice the use of a solvent that can be separated from water is preferred, and considering their application to foods, the use of hexane is preferred. Evaporation of organic solvents from the extract under reduced pressure yields a lipid containing unsaturated fatty acids.

In the lipids obtained as above, unsaturated fatty acids are present in the form bound to triglyceride, phosphatidylcholine, phosphatidylethanolamine, phosphatidylinositol and the like. The lipids are composed of glycerides (triglycerides, diglycerides, monoglycerides), phospholipids, fatty acids, glycolipids,

sterol esters and the like, and glycerides in the lipid are 50 to 100%, preferably 70 to 100%, phospholipids are 0-50%, preferably 0 to 30%, fatty acids, glycolipids, and sterol esters combined are 0 to 30%, preferably 0 to 15%.
5 Triglycerides in the lipids are 50 to 100%, preferably 70 to 100%.

The lipid composition in a lipid containing unsaturated fatty acids produced using the microorganisms of the present invention derived from microorganisms
10 belonging to genus *Mortierella* is 70 to 100% by weight of neutral lipids, 0 to 30% by weight of polar lipids, and triglyceride, a main component of the neutral lipids, is 70 to 99% by weight in the lipids. The contents of unsaturated fatty acids may vary depending on the
15 microorganisms and culture conditions used, and the content of arachidonic acid relative to the total fatty acids is not smaller than 10% by weight, preferably 20 to 100% by weight, and more preferably 40 to 100% by weight. The ratio of arachidonic acid to the total fatty acids in
20 triglycerides is not smaller than 10% by weight, preferably 15 to 100% by weight, and more preferably 35 to 90% by weight. However, the present invention is not limited to lipid vesicles secreted by microorganisms belonging to subgenus *Mortierella*, and fatty acids
25 produced are not limited to arachidonic acid, either. Furthermore, after allowing the cell to incorporate the lipid added to the culture medium, they may be allowed to be secreted as lipids in lipid vesicles, and thereby the lipid composition of lipids in lipid vesicles and the
30 ratio of fatty acids of interest are varied, and in a sense can be freely designed.

In order to isolate and purify triglycerides containing unsaturated fatty acids from an unsaturated fatty acid-containing lipid collected from the lipid
35 vesicles or the culture liquid, standard methods are used such as deoxygenation, degumming, dehydration, steam distillation, molecular distillation, cooling separation,

and column chromatography.

In order to separate unsaturated fatty acids from a lipid containing unsaturated fatty acids, they are used in the form of mixed fatty acids or mixed fatty acid esters and concentrated and separated by conventional methods such as urea addition, cooling separation, and column chromatography.

The lipid vesicles of the present invention contain, in abundance, unsaturated fatty acids in the form of triglycerides. Their applications include, but not limited to, raw materials of foods, beverages, cosmetics, pharmaceuticals, animal feeds and the like and additives thereof. Besides, their objective of uses and the amount used are not limited in any way.

For example, foods include, in addition to general foods, functional foods, nutrient supplements, formula for premature infants, formula for babies, baby foods, foods for pregnant women or foods for the aged people. The lipid vesicles of the present invention have excellent dispersion properties to water or hydrophilic substances, and therefore they can be added to foods containing no fat and oil the addition to which having conventionally been impossible, in particular various beverages. Furthermore, since the lipid vesicles of the present invention can stably retain lipids against air oxidation, the addition to foods containing conventional fat and oil can be effected more easily. As foods containing fat and oil, there can be mentioned, for example, natural foods originally containing fat and oil such as meat, fish, or nuts, foods such as soup to which fat and oil are added during cooking, processed foods that uses fat and oil as heat medium such as doughnuts, fat and oil foods such as butter, processed foods to which fat and oil are added during processing such as cookies, or foods to which fat and oil are sprayed or pasted during processing finishing such as hard biscuits. Furthermore, they may be in the form of functional foods

and pharmaceuticals, and for example they may be in the processed form such as enteric nutrients, powders, granules, troches, medicines, suspensions, emulsions, syrups, and drinks.

5

Examples

The present invention will now be explained in more details with reference to specific examples. It should be noted, however, that the present invention will not
10 limited by these examples in any way.

Example 1. Obtaining a microbial strain that
extracellularly secretes lipid vesicles by
mutation of Mortierella alpina IF08568

Mortierella alpina IF08568 was inoculated into a
15 large slant bottle containing 300 ml of Czapek agar medium (0.2% NaNO_3 , 0.1% K_2HPO_4 , 0.05% $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.05% KCl , 0.01% $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, 3% sucrose, 2% agar, pH 6.0), and was cultured at 28°C for 2 weeks.

After culturing, 50 ml of sterile water to which had
20 been added 2 drops of Tween 80 was added to the large slant bottle, which was shaken sufficiently, and then filtered with 4 ply gauze. This procedure was repeated twice, and the filtrate was centrifuged at 8000 x g for 10 minutes. Spores thus obtained were suspended into 50
25 mM Tris/maleate buffer solution (pH 7.5) to 1×10^6 /ml to prepare a spore solution.

To 1.0 ml of the spore solution thus obtained, 0.5 ml of 100 mM Tris/maleate buffer solution (pH 7.5) was added, and 500 μl of the NTG solution (5 mg of N-methyl-
30 N'-nitro-N-nitrosoguanidine per ml of deionized water) was added, which was subjected to mutation treatment by incubating at 28°C for 15 minutes. After adding 3 ml of 10% $\text{Na}_2\text{S}_2\text{O}_3$, the reaction mixture was centrifuged at 5500 x g for 10 minutes, and the precipitate (spores subjected
35 to mutation treatment) was washed with 3 ml of sterile water and centrifuged at 5500 x g for 10 minutes, to

which sterile water was added to prepare a NTG-treated spore suspension.

5 The NTG-treated spore suspension was diluted to about 10^{-3} to 10^{-4} and then plated on a GY agar plate (1% glucose, 0.5% yeast extract, 0.005% Triton X-100, 1.5% agar, pH 6.0). After incubating at 28°C, those that developed colonies were examined for morphology with a result that microbial strains having a growth morphology distinctly different from that of the parent strain were
10 obtained. The entire colonies of the highly unsaturated fatty acid-producing microbial strains including the parent strain were covered with mycelia as the strain accumulate the produced lipids in the cell, whereas the mutants obtained were covered with lipid vesicles.

15 Subsequently, 4 ml of a transparent liquid medium (4% glucose, 1% yeast extract, pH 6.0) was dispensed into a test tube and sterilized at 120°C for 20 minutes. Then a platinum loopful of the microbial strain obtained above was inoculated thereinto and cultured under shaking at
20 28°C for 2 days, which made the medium cloudy. Even after allowing the medium to stand for over 10 minutes, no lipid layers were observed on the surface of the medium. Thus, the lipid was possibly secreted as lipid vesicles.

25 The lipid vesicles that covered the colonies obtained in the culturing on the GY agar plate were analyzed for lipid by thin layer chromatography (TLC). To a previously activated plate (Merck 5554, 200 x 200 x 0.25 mm, silica gel 60F-254, aluminium sheet), the sample
30 and the control (phospholipids, triglycerides, fatty acids) were plated, which was developed with n-hexane : diethylether : acetic acid = 80 : 20 : 2 (V/V/V) using phosphomolybdic acid (10% phosphomolybdic acid in ethanol) and primulin (0.01% primulin in 80% acetone) as
35 a color developer. For primulin, bands were examined under UV light of a long wavelength (366 nm). As a result, the majority of the lipids in the lipid vesicles

observed outside of the cell were found to be triglycerides.

Thus, about 3000 colonies yielded mutants *Mortierella alpina* SAM2241 FERM BP-7272 and SAM2242 that extracellularly secrete lipid vesicles encapsulating mainly triglycerides having highly unsaturated fatty acids as constituent fatty acids.

Example 2. Fatty acid analysis of an extracellularly secreted lipid when *Mortierella alpina* SAM2241 that extracellularly secretes lipid vesicles was cultured on various media

Four ml each of medium A, B, C, D, E, and F was distributed in a test tube, and was sterilized at 120°C for 20 minutes. A platinum loopful of *Mortierella alpina* SAM2241 (FERM BP-7272) obtained in Example 1 was inoculated into the medium, and cultured under shaking at 28°C for 2 days and then at 12°C for 7 days. After culturing, the cells and the filtrate were separated by filtration. The filtrate obtained was placed in a screw-capped test tube (16.5 mmφ), and lyophilized. To this were added 1 ml of methylene chloride and 2 ml of anhydrous methanol-hydrochloric acid (10%), which was methylesterified by treating at 50°C for 3 hours. Four ml of n-hexane and 1 ml of water were added to this, and then extracted twice. The solvent after extraction was evaporated using a centrifuge evaporator (40°C, 1 hour), and the fatty acid methylesters thus obtained were analyzed by capillary gas chromatography. At the time of adding the methylester, 0.2 mg/ml n-heptadecanoic acid (17:0) was added as an internal standard, and fatty acids were quantitated based on the ratio of surface area of GLC.

Medium A	Glucose	1.0%
	K ₂ HPO ₄	0.3
	MgSO ₄ ·7H ₂ O	0.02

	Polypeptone	1.5
	NaCl	0.2
	Yeast extract	0.1
	pH 7.0	
Medium B	Polypeptone	1.0%
	Meat extract	0.5
	Yeast extract	0.1
	NaCl	0.5
	pH 7.0	
Medium C	Glucose	5.0%
	Polypeptone	0.5
	KH ₂ PO ₄	0.2
	K ₂ HPO ₄	0.1
	MgSO ₄ ·7H ₂ O	0.02
	Yeast extract	0.1
	pH 6.5	
Medium D	Glucose	2.0%
	Yeast extract	1.0
	Bactopeptone	1.0
Medium E	Glucose	2.0%
(MRS medium)	Meat extract	1.0
	Yeast extract	0.5
	Casein trypsin digest	1.0
	K ₂ HPO ₄	0.2
	Sodium acetate	0.5
	Diammonium citrate	0.2
	MgSO ₄ ·7H ₂ O	0.02
	MnSO ₄ ·7H ₂ O	0.02
	Tween 80	0.1
Medium F	Glucose	1.0%
	Yeast extract	0.5
	pH 6.5	

The result is shown in Table 1.

Table 1 The composition of extracellularly secreted fatty acids in the lipid obtained on various media

	16:0	18:0	18:1 (n-9)	18:2 (n-6)	18:3 (n-6)	DGLA	AA	Others
Medium A	14	8	20	11	4	4	31	8
Medium B	15	6	28	16	9	-	26	-
Medium C	17	14	13	10	3	2	37	4
Medium D	14	9	23	9	5	2	31	7
Medium E	16	3	30	11	7	2	27	4
Medium F	15	5	23	10	5	3	32	7

16:0, palmitic acid; 18:0, stearic acid; 18:1 (n-9),
5 oleic acid; 18:2 (n-6), linoleic acid; 18:3 (n-6), γ -linolenic acid; DGLA, dihomog γ -linolenic acid; AA, arachidonic acid

10 In any of the media, a lipid containing unsaturated fatty acids were observed to be extracellularly secreted. Furthermore, the total amount of the extracellularly secreted lipid obtained for medium A to F and the amount of arachidonic acid were found to be positively
15 correlated with glucose concentration. Thus, the total amount of lipid per test tube was 0.18 and 0.6 mg at a glucose concentration of 1%, 0.53 and 0.96 mg at a glucose concentration of 2%, and 2.11 mg at a glucose concentration of 5%, and the amount of arachidonic acid per test tube was 0.05 and 0.15 mg at a glucose
20 concentration of 1%, 0.11 and 0.19 mg at a glucose concentration of 2%, and 0.62 mg at a glucose concentration of 5%.

A result with similar tendency was obtained for the mutant SAM2242.

25 Example 3. The amount of arachidonic acid produced by aerated agitating culture using a 10 L jar fermentor of *Mortierella alpina* SAM2241 that extracellularly secretes lipid vesicles

Five liters of a medium (A: pH 5.0, B: pH 6.0, C: pH

7.0) containing 2% glucose, 1.5 % soy flour, 0.3% KH_2PO_4 , 0.1% Na_2SO_4 , 0.05% $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, 0.05% $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, and 0.2% soybean oil was placed in a 10 L jar fermentor, and sterilized at 120°C for 30 minutes. *Mortierella alpina* SAM2241 (FERM BP-7272) obtained in Example 1 was inoculated therein, and were subjected to aerated agitating culture at an aeration rate of 1.0 vvm and a culture temperature of 24°C for 10 days. 2.0% glucose was added on day 1 of culturing, 1.5% glucose on day 2, 1.0% glucose on days 3 and 4, and 0.5% glucose on days 5 and 6.

Sampling was carried out every day. The culture liquid was separated by filtration into the cells and the filtrate (extracellularly secreted lipid vesicles are dispersed therein). The cells were dried at 105°C for 2 hours, and 20 mg of the dried cells was placed in a screw-capped test tube (16.5 mmφ) and was subjected to methylesterification as in Example 2. The filtrate (1 ml) was placed in a screw-capped test tube (16.5 mmφ), was lyophilized, and then was subjected to methylesterification as in Example 2. The fatty acid esters thus obtained were analyzed by capillary gas chromatography. Table 2 shows the amount produced of arachidonic acid and its content on day 9 of culturing.

Table 2 The amount produced of arachidonic acid and its content on day 9 of culturing

		In the cell	Outside of the cell	Extracellular arachidonic acid percentage (%)
Medium A	pH 5.0	6.4g/L(33.2%)	0.14g/L(37.4%)	2.1
Medium B	pH 6.0	5.8g/L(32.4%)	0.24g/L(34.5%)	4.0
Medium C	pH 7.0	3.6g/L(29.7%)	0.43g/L(30.8%)	10.7

Figures in parentheses indicate the ratio of arachidonic acid relative to the total fatty acids.

With increased pH of the medium, the extracellular

secretion of arachidonic acid-containing lipids was promoted.

Example 4. Lipid analysis of lipid vesicles
secreted by *Mortierella alpina* SAM2241 that
extracellularly secretes lipid vesicles

The culture filtrates on day 9 of culturing in medium A, B, and C obtained in Example 3 were treated with chloroform/methanol/water (1:2:0.8) by the Bligh-Dyer method and the total lipids were extracted from the extracellularly secreted lipid vesicles. The total lipids obtained contained neutral lipids (triglycerides) and polar lipids (phospholipids). The total extracted lipids were charged into the Sep-pak Silica cartridge (manufactured by Waters) and eluted to obtain the neutral lipid fraction with chloroform and the polar lipid fraction with methanol. After evaporating the solvent, methylesterification was carried out as in Example 2, and fatty acid methylesters obtained were analyzed by capillary gas chromatography. As a percentage of the total fatty acids to which triglycerides and phospholipids bind, the percentages of triglyceride and phospholipids were calculated. As a result, the percentage of the triglycerides in the total lipids for medium A, B, and C were 95.3%, 97.7%, and 96.2%, respectively.

Example 5. Continuous culture of *Mortierella alpina*
SAM2241 that extracellularly secretes lipid
vesicles in a 10 L bioreactor

To a 10 L bioreactor having two built-in ceramic filters, 5 L of a medium containing 2% glucose and 2% yeast extract with pH adjusted to 7 was prepared, to which a precultured microbial strain of *Mortierella alpina* SAM2241 (FERM BP-7272) obtained in Example 1 was inoculated and subjected to aerated agitating culture. On the next day, a glucose solution was added through a ceramic filter to increase the glucose concentration in the medium by 3%. On day 2 also, the glucose solution

was added through the ceramic filter to increase the glucose concentration in the medium by 3%.

On day 3 and after, a 5% glucose solution and a 0.05% yeast extract solution were continuously passed through a ceramic filter at a speed of about 1000 ml/day. And the culture liquid was continuously extracted through a ceramic filter at about 600 ml/day (the amount of liquid is adjusted to remain constant). In order to prevent the clogging of the filter with the cells, feeding of the glucose and yeast extract solutions and extracting of the culture liquid were alternately carried out as appropriate. Due to evaporation of water vapor by aeration, the amount of liquid in the jar remained almost constant. The feeding speed of glucose was adjusted by the glucose concentration to be extracted.

As a result, a medium (culture liquid) that contained about 1 g/L arachidonic acid-containing triglycerides was able to be continuously extracted.

Example 6. Microbial transformation of the fat and oil added to the medium and its migration into lipid vesicles by *Mortierella alpina* SAM2241 that extracellularly secretes lipid vesicles

To 2 ml of a medium (pH 6.0) containing 1% glucose and 1% yeast extract, 2% linseed oil or fish oil was added, which was then put into a 10 ml Erlenmeyer flask and sterilized at 120°C for 20 minutes. One platinum loopful of *Mortierella alpina* SAM2241 (FERM BP-7272) obtained in Example 1 was inoculated into the medium, and cultured using a reciprocating shaker (150 rpm) at 28°C for 8 days. The filtrate was recovered by filtration, was lyophilized, and then the extracellularly secreted lipid was subjected to methylesterification as in Example 2, and the fatty acid methylester was analyzed by capillary gas chromatography.

When linseed oil was added to the medium, a major fatty acid of linseed oil, 9,12,15-octadecatrienoic acid

(α -linolenic acid) served as a substrate of the fatty acid biosynthetic enzymes of the mutant, and was converted to 6,9,12,15-octadecatetraenoic acid (stearidonic acid), 8,11,14,17-eicosatetraenoic acid, and 5,8,11,14,17-eicosapentaenoic acid, and the lipid in the lipid vesicles contained 2.4, 3.3, and 8.1% of 6,9,12,15-octadecatetraenoic acid (stearic acid), 8,11,14,17-eicosatetraenoic acid, and 5,8,11,14,17-eicosapentaenoic acid, respectively, confirming that the converted fatty acids are extracellularly secreted as the constituent fatty acids of triglycerides. When fish oil was added to the medium, the fact that 5,8,11,14,17-eicosapentaenoic acid and 4,7,10,13,16,19-docosahexaenoic acid of fish oil are incorporated into the microbial strain and are extracellularly secreted as constituent fatty acids of triglycerides was confirmed because the lipid of the extracellularly secreted lipid vesicles contained 8.1 and 12.2% of 5,8,11,14,17-eicosapentaenoic acid and 4,7,10,13,16,19-docosahexaenoic acid, respectively.

Example 7. Component analysis of lipid vesicles extracellularly secreted by *Mortierella alpina* SAM2241

In order to analyze components of extracellularly secreted lipid vesicles, a spore suspension of *Mortierella alpina* SAM2241 obtained in Example 1 was plated to the GY agar plate (1% glucose, 0.5% yeast extract, 0.005% Triton X-100, 1.5% agar, pH 6.0), and cultured at 28°C for 4 days. As shown in Example 1, the entire colonies were covered with lipid vesicles. Thus, the small vesicles were collected into a screw-capped test tube (16.5 mm ϕ). Chloroform (2 ml) and KCl solution (2 ml) were added thereto, shaken and extracted. Lipids migrated into the chloroform layer and sugars and proteins migrated into the KCl layer, and these were analyzed for the components according to a standard method and were found to comprise sugars: 38.1%,

proteins: 18.2%, lipids: 43.7%.

Example 8. Preparation of formula using lipid vesicles

The culture filtrate obtained in Example 3 was separated using a centrifuge (TOMY RL-101) at 1500 x g and washed with sterile water to prepare lipid vesicles fit for consumption. The lipid vesicles (0.92 g) were added to 100 g of powdered milk to prepare a formula containing lipid vesicles. The composition of arachidonic acid in the formula obtained was 0.5% of the total fatty acids, which was similar to that of the mother's milk.

When the formula was dissolved in water, its dispersion in water was good and uniformly dispersed without any separation of oils.

Example 9. Preparation of capsules

Water was added to 100 parts per weight of gelatin and 35 parts per weight of food additive glycerin, which was dissolved at 50 to 60°C to prepare a gelatin coating with a viscosity of 20000 cps. Then, from the lipid vesicles separated by centrifugation from the culture filtrate obtained in Example 3, lipids were extracted and purified according to a standard method. Then, 97% the refined oil and 3% vitamin E oil were mixed to prepare a content. Using these, capsule molding and drying were carried out according to a standard method so that soft capsules containing 180 mg content per capsule were produced.

Example 10. Preparation of lipid vesicles-containing beverages

The lipid vesicles (10 g) fit for consumption obtained in the method shown in Example 8 were added to 10 L of orange juice to prepare juice containing lipid vesicles.

Reference to the microorganisms deposited under the Patent Cooperation Treaty, Rule 13-2, and the name of the Depository Authority
Depository Authority:

Name: the National Institute of Bioscience and Human
Technology, Agency of Industrial Science and Technology
Address: 1-3, Higashi 1-chome, Tsukuba city, Ibaraki
Pref., Japan

5 Organism (1)

Name: *Mortierella elongata* SAM0219

Accession number: FERM BP-1239

Date deposited: March 19, 1986

Organism (2)

10 Name: *Mortierella alpina* SAM2241

Accession number: FERM BP-7272

Date deposited: August 11, 2000

CLAIMS

1. A microorganism which extracellularly secretes an unsaturated fatty acid-containing lipid as lipid vesicles encapsulating said lipid.

5 2. The microorganism according to claim 1 wherein said unsaturated fatty acids are fatty acids having 18 carbons and two or more double bonds.

3. The microorganism according to claim 1 or 2 which is a filamentous fungus.

10 4. The microorganism according to claim 3 which is a microorganism belonging to genus *Mortierella*.

5. The microorganism according to claim 4 which is a microorganism belonging to genus *Mortierella* subgenus *Mortierella*.

15 6. The microorganism according to claim 5 which is a microorganism belonging to the species *alpina*.

7. The microorganism according to any one of claims 1 to 6 which has a property of forming lipid vesicles containing a lipid around the colonies when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

20 8. The microorganism according to any one of claims 1 to 7 which is obtained by artificially treating a microorganism having an ability to accumulate an unsaturated fatty acid-containing lipid in the cell.

25 9. The microorganism according to any one of claims 1 to 7 which is selected by artificially treating a microorganism having an ability of accumulating an unsaturated fatty acid-containing lipid in the cell, by culturing the obtained strains on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and then by selecting those strains that make the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

30 10. The microorganism according to any one of

claims 1 to 9 which can be turned into a spheroplast or a protoplast.

5 11. A filamentous fungus having a property of extracellularly secreting a lipid containing unsaturated fatty acids.

12. The microorganism according to any one of claims 1 to 11 wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

10 13. The microorganism according to any one of claims 1 to 12 wherein said unsaturated fatty acids are arachidonic acid.

14. The microorganism according to claim 13 wherein said lipid contains 10% or more arachidonic acid relative to the total fatty acids.

15 15. Lipid vesicles encapsulating an unsaturated fatty acid-containing lipid.

20 16. The lipid vesicles according to claim 15 wherein said unsaturated fatty acids are unsaturated fatty acids having 18 or more carbons and two or more double bonds.

17. The lipid vesicles according to claim 15 or 16 wherein said lipid vesicles are produced by a microorganism.

25 18. Lipid vesicles encapsulating a lipid obtained from a culture liquid prepared by culturing the microorganism according to any one of claims 1 to 14 in a liquid medium.

30 19. The lipid vesicles according to any one of claims 15 to 18 which can be uniformly dispersed in water or a hydrophilic substance.

20. The lipid vesicles according to any one of claims 15 to 18 which stably retains the lipid encapsulated within said lipid vesicles against oxidation.

35 21. The lipid vesicles according to any one of claims 15 to 18 which can be separated by centrifugation.

22. The lipid vesicles according to any one of

claims 15 to 21 wherein the membrane of said lipid vesicles comprises sugar, protein, and lipid.

23. The lipid vesicles according to any one of claims 15 to 22 which has an average diameter of 0.2 to 10 μ m.

24. The lipid vesicles according to any one of claims 15 to 23 wherein the lipid encapsulated in said lipid vesicles is a lipid in which 50% or more is triglyceride.

25. A lipid isolated from the lipid vesicles according to any one of claims 15 to 24.

26. A food, a cosmetic, or an animal feed comprising the lipid vesicles according to any one of claims 15 to 24 added thereto.

27. The food according to claim 26 wherein the food comprising the lipid vesicles added thereto is a functional food, a nutrient supplement, formula for premature infants, modified milk for babies, a baby food, a food for pregnant women or a food for the aged people.

28. The food according to claim 26 wherein the foods to which the lipid vesicles have been added are beverages.

29. A food, a cosmetic, a pharmaceutical or an animal feed comprising the lipid according to claim 25 added thereto.

30. A method of producing lipid vesicles which method comprises culturing the microorganism according to any one of claims 1 to 14 in a liquid medium and then collecting the lipid vesicles encapsulating a lipid from the culture liquid.

31. A method of producing lipid vesicles which method comprises continuously culturing the microorganism according to any one of claims 1 to 14 in a liquid medium and then continuously collecting the lipid vesicles encapsulating a lipid from the culture liquid.

32. A method of producing a lipid which method comprises culturing the microorganism according to any

one of claims 1 to 14 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, and separating a lipid containing fatty acids from said lipid vesicles.

5 33. A method of producing unsaturated fatty acids which method comprises culturing the microorganism according to any one of claims 1 to 14 in a liquid medium, collecting lipid vesicles encapsulating a lipid from the culture liquid, separating the lipid containing
10 fatty acids from said lipid vesicles, and isolating the unsaturated fatty acids from said lipid.

 34. A microorganism having a property of extracellularly secreting a lipid containing unsaturated fatty acids that have 18 carbons and three or more double
15 bonds or 20 or more carbons and two or more double bonds.

 35. A microorganism having a property of directly and extracellularly secreting a lipid containing unsaturated fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more
20 double bonds.

 36. The microorganism according to claim 34 or 35 which is a filamentous fungus.

 37. The microorganism according to any one of claims 34 to 36 which has a property of forming lipid-
25 containing lipid vesicles around the colonies thereof when said microorganism is grown on a solid medium, and/or of making the culture liquid cloudy when said microorganism is cultured in a transparent liquid medium.

 38. The microorganism according to any one of
30 claims 34 to 37 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

35 39. The microorganism according to claim 35 or 36 obtained by artificially treating a microorganism which has an ability of intracellularly accumulating a lipid

5 containing fatty acids that have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds, and by selecting, from the strains obtained, strains that make the culture liquid cloudy and then separates a lipid layer when cultured in a transparent liquid medium.

40. The microorganism according to any one of claims 34 to 39 which can be turned into a spheroplast or a protoplast.

10 41. The microorganism according to any one of claims 34 to 40 wherein said extracellularly secreted lipid is a lipid in which 50% or more is triglyceride.

15 42. A method of producing a lipid containing unsaturated fatty acids which method comprises culturing the microorganism according to any one of claims 34 to 41 in a liquid medium and collecting the lipid from the culture liquid.

20 43. A method of producing a lipid containing unsaturated fatty acids which method comprises continuously culturing the microorganism according to any one of claims 34 to 41 in a liquid medium and then continuously collecting the lipid from the culture liquid.

25 44. A screening method wherein a microorganism having an ability of extracellularly secreting a lipid containing unsaturated fatty acids using as an index the fact that when the microorganism is cultured using a transparent liquid medium the culture liquid becomes cloudy.

30 45. The screening method according to claim 44 wherein said unsaturated fatty acids have 18 carbons and three or more double bonds or 20 or more carbons and two or more double bonds.

35 46. The screening method according to claim 44 wherein said microorganism is a filamentous fungus.

47. A screening method wherein strains having a property of extracellularly secreting an unsaturated

5 fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, and by culturing the strains obtained on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery.

10 48. A screening method wherein strains having a property of extracellularly secreting an unsaturated fatty acid-containing lipid are selected by artificially treating a microorganism having an ability to accumulate the unsaturated fatty acid-containing lipid in the cell, by culturing the strains obtained on a solid medium to select strains of which colonies are covered with lipid-containing lipid vesicles at the periphery, and by
15 further culturing the selected strains in a transparent liquid medium to select strains for which the culture liquid becomes cloudy.

20 49. The screening method according to claim 47 or 48 wherein said artificial manipulation is mutation treatment with N-methyl-N'-nitro-N-nitrosoguanidine (NTG).

50. The screening method according to claim 47 or 48 wherein said artificial manipulation is mutation treatment, gene manipulation, or cell fusion.

25 51. A microorganism selected by the screening method according to any one of claims 44 to 50.

ABSTRACT

There are provided microorganisms having a property of producing a lipid containing unsaturated fatty acids as constituent fatty acids and extracellularly secreting the produced lipid encapsulated in lipid vesicles, methods of screening said microorganisms, as well as methods of efficiently producing a fatty acid-containing lipid using said microorganisms. Furthermore, there are provided lipid vesicles encapsulating a lipid containing unsaturated fatty acids, and foods, cosmetics, and animal feeds comprising said lipid vesicles added thereto.

Artificially treated microorganisms or microorganisms collected from nature are grown on a solid medium, and microbial strains that form lipid vesicles at the periphery of the colonies and/or microbial strains that, when cultured in a transparent liquid medium, make the culture liquid cloudy are selected. The microorganisms obtained are cultured, lipid-containing lipid vesicles secreted in the culture liquid, are separated from the culture liquid, and the lipid is separated and purified.

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Declaration and Power of Attorney For Patent Application

特許出願宣言書及び委任状

Japanese Language Declaration

日本語宣言書

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My residence, post office address and citizenship are as stated next to my name.

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I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

MICROORGANISMS THAT EXTRACELLULARLY
SECRETE LIPIDS AND METHODS OF PRODUCING
LIPID AND LIPID VESICLES ENCAPSULATING
LIPIDS USING SAID MICROORGANISMS

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PCT/JP00/05425 and was amended on
___ (if applicable).

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I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

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Prior Foreign Application(s)

外国での先行出願
11-229509(Pat. Appln.)

(Number)
(番号)

Japan

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(国名)

(Number)
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I hereby claim foreign priority under Title 35, United States Code, Section 119 (a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Priority Not Claimed

優先権主張なし

13/August/1999

(Day/Month/Year Filed)
(出願年月日)

(Day/Month/Year Filed)
(出願年月日)

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(Application No.)
(出願番号)

(Filing Date)
(出願日)

I hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s), or 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code Section 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.86 which became available between the filing date of the prior application and the national or PCT International filing date of application.

(Status: Patented, Pending, Abandoned)
(現況: 特許許可済、係属中、放棄済)

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Japanese Language Declaration (日本語宣言書)

委任状: 私は下記の発明者として、本出願に関する一切の手続きを米特許商標局に対して遂行する弁理士または代理人として、下記の者を指名いたします。(弁理士、または代理人の氏名及び登録番号を明記のこと)

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith (list name and registration number)

William L. Mathis 17,337
Peter H. Smolka 15,913
Robert S. Swecker 19,885
Platon N. Mandros 22,124
Benton S. Duffett, Jr. 22,030
Joseph R. Magnone 24,239
Norman H. Stepmo 22,716
Ronald L. Grudziecki 24,970
Frederick G. Michaud, Jr. 26,003
Alan E. Kopecki 25,813
Regis B. Slutter 26,999
Samuel C. Miller, III 27,360
Ralph L. Freeland, Jr. 16,110

Robert G. Mukai 28,531
George A. Hovance, Jr. 28,223
James A. LaBarre 28,632
E. Joseph Gess 28,510
R. Danny Huntington 27,903
Eric H. Welsblatt 30,505
James W. Peterson 26,057
Teresa Stanek Rea 30,427
Robert E. Krebs 25,885
William C. Rowland 30,888
T. Gene Dillahunty 25,423
Patrick C. Keane 32,858
Bruce J. Boggs, Jr. 32,344

William H. Benz 25,952
Peter K. Skiff 31,917
Richard J. McGrath 29,195
Matthew L. Schneider 32,814
Michael G. Savage 32,596
Gerald F. Swiss 30,113
Michael J. Ure 33,089
Charles F. Wieland III 33,096
Bruce T. Wieder 33,815
Todd R. Walters 34,040

書類送付先

Send Correspondence to:

Ronald L. Grudziecki
BURNS, DOANE, SWECKER & MATHIS, L.L.P.
P.O. Box 1404
Alexandria, Virginia 22313-1404

直接電話連絡先: (名前及び電話番号)

Direct Telephone Calls to: (name and telephone number)

Ronald L. Grudziecki
at (703) 836-6620

第一または第一発明者名

Full name of sole or first inventor

Kengo Akimoto

発明者の署名

日付

Inventor's signature

Date

Kengo Akimoto

April 5, 2001

住所

Residence

Mishima-gun, Osaka, Japan

国籍

Citizenship

Japanese

私書箱

Post Office Address

1-9-5-1006, Yamazaki, Shimamoto-cho,

Mishima-gun, Osaka 618-0001, Japan

第二共同発明者

Full name of second joint inventor, if any

Hiroshi Kawashima

第二共同発明者

日付

Second inventor's signature

Date

Hiroshi Kawashima

April 5, 2001

住所

Residence

Takatsuki-shi, Osaka, Japan

国籍

Citizenship

Japanese

私書箱

Post Office Address

6-11-1-113, Magami-cho, Takatsuki-shi,

Osaka 569-1121, Japan

(第三以降の共同発明者についても同様に記載し、署名をすること)

(Supply similar information and signature for third and subsequent joint inventors.)

第三共同発明者

Full name of third joint inventor, if any
Sakayu Shimizu

第三共同発明者

日付

Third inventor's signature

Date

Sakayu Shimizu

April 5, 2001

住 所

Residence

Kyoto-shi, Kyoto, Japan

国 籍

Citizenship

Japanese

私書箱

Post Office Address

6-9, Tokiwayamashita-cho, Ukyo-ku, Kyoto-shi,

Kyoto 616-8212, Japan

第四共同発明者

Full name of fourth joint inventor, if any

第四共同発明者

日付

Fourth inventor's signature

Date

住 所

Residence

国 籍

Citizenship

私書箱

Post Office Address

第五共同発明者

Full name of fifth joint inventor, if any

第五共同発明者

日付

Fifth inventor's signature

Date

住 所

Residence

国 籍

Citizenship

私書箱

Post Office Address

第六共同発明者

Full name of sixth joint inventor, if any

第六共同発明者

日付

Sixth inventor's signature

Date

住 所

Residence

国 籍

Citizenship

私書箱

Post Office Address

(第七以降の共同発明者についても同様に
記載し、署名をすること)

(Supply similar information and signature for
seventh and subsequent joint inventors.)